ORIGINAL RESEARCH



Tautomerism of 4-phenyl-2,4-dioxobutanoic acid. Insights from pH ramping NMR study and quantum chemical calculations

Ilija N. Cvijetić¹ • Miloš P. Pešić² • Miljana D. Todorov² • Branko J. Drakulić³ • Ivan O. Juranić³ • Tatjana Ž. Verbić² • Mire Zloh⁴

Received: 28 July 2017 / Accepted: 19 September 2017 © Springer Science+Business Media, LLC 2017

Abstract Aryldiketo acids (ADKs) exhibit the variety of biological activities, mainly due to large affinity toward divalent metal ions. Metal complexation ability of ADKs, as well as interactions with proteins, depend on tautomeric form present in solution. The main aim of this study was to fully explore the tautomeric preferences of 4-phenyl-2,4-dioxobutanoic acid (4PDA), as ADKs representative, in aqueous media at different pH values. 1D and 2D NMR spectroscopy in combination with quantum chemical calculations was applied in order to better understand the tautomeric preferences of 4PDA. The data in highly acidic media are especially interesting since there are no such findings in the literature due to low solubility

We regret to inform that Branko Drakulić has passed away since completion of this work.

Electronic supplementary material The online version of this article (https://doi.org/10.1007/s11224-017-1039-3) contains supplementary material, which is available to authorized users.

Ilija N. Cvijetić ilija@chem.bg.ac.rs

- Tatjana Ž. Verbić tatjanad@chem.bg.ac.rs
- ¹ Innovation Center of the Faculty of Chemistry, Ltd, Studentski Trg 12-16, Belgrade 11000, Serbia
- ² Faculty of Chemistry, University of Belgrade, Studentski Trg 12-16, Belgrade 11000, Serbia
- ³ Department of Chemistry-IChTM, University of Belgrade, Njegoševa 12, Belgrade 11000, Serbia
- ⁴ School of Life and Medical Sciences, University of Hertfordshire, Hatfield AL10 9AB, UK

of ADKs in molecular form. At low pH values, where 4PDA is unionized, the most abundant tautomeric form is enol with keto group closer to phenyl ring. At higher pH values, mixture of two 4PDA ionic forms coexists in solution. Their ratio calculated according to NMR data fits the values predicted using two experimentally determined pK_a values. Based on the complexity of ¹H NMR spectrum of monoanionic 4PDA form, coexistence of two stable rotamers was assumed. In an alkaline media, 4PDA is mostly present in dianionic form. As π -electrons of dianion are delocalized over an entire keto-enol moiety, spectral distinction between tautomers was not possible. Quantum chemical calculations were used to predict relative stability of tautomers. The predictions were in good accordance with experimental results only in case when explicit water molecule was included in calculations.

Keywords Aryldiketo acid · Keto-enol tautomerism · NMR spectroscopy · Quantum chemical calculations

Introduction

Molecular properties and biological activities of aryldiketo acids (ADKs), an important class of molecules with widespread biological activities [1–6], are affected by keto-enol tautomerism. Tautomerism is of particular interest in studies of small organic molecules recognition properties, including protein-ligand interactions, since different tautomers of molecules of this class have different hydrogen-bond acceptor/ hydrogen-bond donor patterns, as well as different metal complexation abilities [7].

ADKs act by functional sequestration of Mg^{2+} ion in the active center of HIV-1 integrase (IN) [8], an enzyme

responsible for the integration of viral DNA into the host genome. It was shown that ADKs' metal complexation ability depends on tautomeric form that is predominant in solution [9], and that Mg²⁺ preferentially reacts with enolate form I (Scheme 1) [10]. Similarly, in crystal structures of influenza virus PA protein (PDB entries 4E5G and 4E5H) and La Crosse orthobunyavirus L-protein (PDB entry 2XI7) ADKs are involved in interactions with two Mn²⁺ ions [11, 12]. In crystal structures of ADKs complexed with various bacterial enzymes, both diketo and enol forms are found [13, 14]. Furthermore, hydrolytic C–C bond cleavage of 1,3-diketones (β -diketones) by β -ketolases (EC 3.7.1.1 to EC 3.7.1.10) is sensitive to tautomeric form in which a 1,3-diketone is present in solution [10, 15, 16].

ADKs simultaneously exist in two enolate forms (Scheme 1, I and III), conformationally locked by the pseudo-ring, and one diketo form (II) having two rotatable bonds responsible for their conformational flexibility [15, 17]. As a continuation of our group's work on structure-property relationships [18–20] and biological activities of ADKs [6], we aimed to further explore their tautomeric preferences.

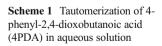
It was shown previously that the enol form of 1,3-diketones is thermodynamically favored compared to diketo form due to stabilization via intramolecular hydrogen bonding. In the ground state of avobenzone (1,3-diketone used in sunscreen products as a UV light absorber), the presence of other forms such as non-chelated enol and its rotamers is also hypothesized [21, 22].

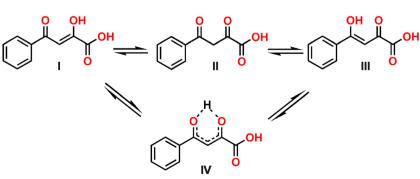
Keto-enol tautomerization of 1,3-diketones has been studied extensively in various chemical systems using a range of analytical techniques (NMR, IR, HPLC, gas electron diffraction) [15, 23, 24]. It is enhanced in polar, protic solvents. Density functional theory (DFT) calculations showed that tautomerization is highly enhanced in water, as two water molecules assist process via transition state analog to E2 mechanism TS [25]. Tautomerization may be difficult without highly activated proton relays of water hydrogen bonds, i.e., the methylene C–H bond in the keto form cannot be cleaved readily by the nucleophilic sources other than a water molecule connected by hydrogen-bond networks. The newly formed enol O–H bond participates in the intermolecular hydrogen bond rather than in the intramolecular one. NMR spectroscopy and DFT calculations of phenindione (cyclic 1,3-diketone) and derivatives showed that the predominant tautomer for these compounds in DMSO solution is enol form [26].

In a quantum chemical study of the structure and stability of diketo acid HIV-1 IN inhibitors, 5-CITEP and L-731,988, enol forms were more stable than diketo form, with the energy differences for these molecules ranging from 15 to 28 kJ/mol [17]. Two enol forms had similar energies with only 3 kJ/mol difference. Because practically no energy barrier between these two enol forms was observed, it was proposed that they can interconvert easily, and a delocalized transition state (sixmembered ring) was suggested.

Detailed study of keto-enol tautomerism of 11 4-alkyl- and 4-aryl-2,4-diketobutanoic acids has shown that enolate I (Scheme 1) is a predominant form (98%) in a protic solvent $(CDCl_3)$ [15]. The equilibrium ratios of aqueous solution structures of aliphatic 2,4-diketobutanoic acids (2,4-diketo, 2-enol-4-keto, and 2-hydrate-4-keto) were markedly affected by solution pH value within wide pH range (1.5–10.5). At pH 7.5 ratio of these structures was approximately 4:5:1, at low pH values 2-hydrate predominated ($\approx 50\%$) and at high pH values 2-enolate carboxylate was dominant ($\approx 80\%$) while 2-hydrate was not detected. As aromatic 2,4-diketobutanoic acids are less soluble in acidic media, a tautomerization study was performed only in solutions with $pH \ge 5.5$. The authors reported that three tautomeric forms (I-III, Scheme 1) were not distinguished in the NMR spectra due to the formation of pseudodienolate form (IV) and fast interconversion between two enolate forms. To the best of our knowledge, no experimental data on keto-enol tautomerism of ADKs in aqueous solution with pH \leq 5.5 was published so far.

In order to fully explore a potential of ADKs as biologically active molecules with metal complexing ability, it is important to have knowledge about their tautomeric forms in aqueous media at different pH values. NMR experiments at a wide range of pH values, starting from extreme acidic to basic conditions, are performed for assessing the tautomeric preferences of 4PDA as a representative scaffold for active ADKs. Results are explained and rationalized with the aid of theoretical calculations and previously determined acid-base properties of the compound of interest.





Experimental

Materials and methods

Synthesis and characterization of 4PDA were already described [20]. The same sample was used in all NMR measurements described herein. All other chemicals were purchased from Fluka, Aldrich, or Merck, having > 98% purity, and were used as received.

¹H, ¹³C, COSY, HMQC, and HMBC NMR spectra of 4PDA in aqueous solution at different pH values were acquired using Bruker Avance 500/125 MHz NMR spectrometer. pH values were measured using Corning 120 pH-meter equipped with Corning Ag/AgCl microelectrode.

1D and 2D NMR spectra of 4PDA

All NMR spectra of 4PDA were acquired at $t = 25 \pm 1$ °C, and constant ionic strength (I = 0.1 M (NaNO₃)). TSP was used as the internal standard for spectra calibration, and chemical shifts (δ) are given in *ppm*. Measured pH values are converted to pD according to relation: pD = pH_{measured} + 0.4 [27]. The sample was dissolved in an appropriate solvent (CF₃COOD for pH < 0, D₂O/CD₃COOD for pH 2.09; acetate buffer-*d*—for pH 4.41; and carbonate buffer for pH 7.80 and 9.20), and spectra were calibrated using TMS as an internal standard.

The 1D ¹H spectrum was acquired using a sweep width of 10,330 Hz and 65,536 data, giving a digital resolution of 0.157 Hz/point and an acquisition time of 3.17 s. Pulse width of 11.50 µs and relaxation delay of 2 s were used for 256 transients.

The 1D ¹³C spectrum was acquired using a sweep width of 30,030 Hz and 131,072 data points giving a digital resolution of 0.229 Hz/point and an acquisition time of 2.18 s. Pulse width of 10 μ s and relaxation delay of 2 s were used for 6000 transients.

2D HMQC (pulse program inv4gpqf) and HMBC (pulse program inv4gplpIrndqf—optimized for coupling constant 10 Hz) spectra were acquired with 1024 points in direct dimensions (sweep width 4882 Hz) and 256 complex points (sweep width 31,440 Hz) giving digital resolutions of 4.76 and 122 Hz/point, respectively. Digital resolution for complex dimension has increased to 30.7 Hz/point after zero filling to 1024 data points.

Solubility studies

Intrinsic solubility (solubility of the molecular form (H₂A)) of 4PDA was determined by the "shake-flask" method in 1 M HCl. An excess of the accurately weighed compound was added to 5.0 mL of 1 M HCl; five samples were prepared. All samples were stirred and thermostated overnight at $t = 25 \pm 1$ °C. After equilibration, samples were filtered,

aliquots appropriately diluted with HCl, and the concentration of 4PDA in saturated solution determined spectrophotometrically at the wavelength of the absorption maximum, $\lambda_{\text{max}} = 308.8$ nm. Conformity with Beer's law had previously been verified.

Computational chemistry studies

The full geometry optimizations of different 4PDA tautomers, in neutral (H₂A) and monoanionic (HA⁻) forms, were performed at the MP2 level of theory, using the 6-31G(d,p) basis set, calculating the force constants at every point (Opt=Calcall), and tightening the cutoff for forces and step size to determine the convergence (Opt=Tight).

The NMR shifts of the three tautomers of molecular form of 4PDA with the carboxyl H in *out* orientation were predicted applying the gauge-independent atomic orbital (GIAO) method with MP2/6-311++G(d,p) by SP calculations on fully optimized geometries [28]. Implicit water solvation model (IEF-PCM) was applied. The GIAO predicted magnetic shielding of tetramethylsilane (TMS) was taken as a reference.

For the optimization of the systems comprising one water molecule and different tautomeric forms of 4PDA, we used optimized geometries of tautomers and manually added one explicit water molecule in the proximity of carboxyl OH. For each tautomer, the system was initially optimized holding 4PDA rigid and allowing movement of the water molecule by semiempirical MO PM6 method in MOPAC2016 [29]; then full optimization without constraints was performed by MP2/6-31G(d,p). The influence of solvent was also simulated applying the implicit (water) solvation model, IEF-PCM, as default in Gaussian09 [30].

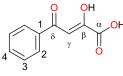
Results and discussion

NMR study of aqueous 4PDA solutions at different pH values

Brecker et al. [15] used the NMR spectroscopy to study the aqueous structures of 4PDA in phosphate buffer at pH 7.5. The authors reported broad signals of C_{γ} H atom, as well as broad signals of C_{β} , C_{γ} , and C_{δ} carbon atoms (for atom labeling see Table 1). Interconversion between two enolic forms is very fast, and therefore, it is impossible to ascribe signals to any distinct tautomer. Signal broadening could be explained by the finding of Guthrie et al. [31] that enolization rate constants of 2,4-diketo acids are significantly increasing as pH value increases.

Maurin et al. [32] studied NMR spectra of 4PDA in pure water, and in buffers pH 7.5 and 10.0. In water, they observed only one species, and attributed signals to fully protonated 4PDA in enol I tautomeric form (Scheme 1). However, as

Table 1 Experimentally obtained and calculated ¹H and ¹³C NMR chemical shifts for three 4PDA tautomeric forms (Scheme 1) in CF₃COOD



Atom	δ (<i>ppm</i>)											
	C_{α}	C_{β}	C_{γ}	$H_{\gamma}\left(H_{\gamma}'\right)$	C_{δ}	C ₁	C ₂	H ₂ (H ₂ ')	C ₃	H ₃ (H ₃ ')	C ₄	H ₄
Experim.	163.41	161.59	95.06	7.23 ^a	189.46	129.85	124.05	7.96 ^b	125.05	7.50 ^c	131.13	7.64 ^d
Enol I calc.	152.58	149.45	85.60	7.70	181.50	126.42	110.75 (111.11)	8.41 (8.74)	116.75 (117.20)	8.07 (8.23)	116.61	7.72
Enol III calc.	151.09	167.25	82.28	7.64	167.28	123.27	109.56 (110.45)	8.22 (8.29)	116.66 (116.88)	8.08 (8.12)	116.23	7.72
Diketo II calc.	150.28	177.19	36.38	3.89 (5.14)	182.29	122.98	110.47 (111.11)	8.12 (8.91)	117.31 (118.49)	8.14 (8.29)	117.49	7.79

^a(*s*, 1H)

 $^{c}(t, J = 7.90 \text{ Hz}, 2\text{H})$

 $^{d}(t, J = 7.47 \text{ Hz}, 1\text{H})$

4PDA in aqueous media act as weak diprotic acid $(pK_{a1} = 2.06; pK_{a2} = 7.56)$ sparingly soluble in water [20], it will partially dissociate, giving a mixture of molecular and monoanionic form. Therefore, we recorded NMR spectra in highly acidic medium (Figs. 1, 2, and 3, Fig. S1) in order to suppress the dissociation and obtain signals of the pure molecular (H₂A) form of 4PDA.

The low intrinsic solubility of 4PDA ($1.08 \pm 0.05 \times 10^{-3}$ M) explains why long (overnight) signal acquisition was necessary for 1D and 2D NMR spectra recording in the highly acidic medium. Full structure-spectra assignments were achieved using COSY, HMQC (ESM, Fig. S1), and HMBC spectra (Fig. 3) of 4PDA recorded in CF₃COOD. Experimental and calculated ¹H and ¹³C NMR chemical shifts of 4PDA are given in Table 1.

Singlet at 7.23 *ppm* in ¹H NMR spectrum (Fig. 1) is the signal of the vinyl group H atom (H_{γ}, Table 1). As the exchange rate between H_{γ} and D atom from CF₃COOD is fast, the integral of this signal is smaller than expected. Furthermore, the lack of a singlet around 4.5 *ppm* in ¹H NMR spectrum (this part of the spectra is not shown in Fig. 1), as well as lack of a signal around 50 *ppm* in ¹³C NMR spectrum (Fig. 2), both expected for > CH₂ group (C_{γ}) in diketo form II, indicate that diketo form does not exist in solution at concentrations detectable by NMR. The presence of hydrate form is also excluded as no signals characteristic for this form were observed. The C_{γ} atom of hydrate form would also give a signal around 50 *ppm* in ¹³C NMR spectrum, as well as the singlet at chemical shift below 4 ppm in ¹H NMR spectrum. This finding is in accordance with findings of

Brecker et al. [15], who found this form in significant amount only in 4-alkyl-2,4-dioxobutanoic acids, but not in 4-phenyl-2,4-dioxobutanoic acid.

Due to H/D exchange, signal at 95 ppm in ¹³C NMR spectrum consists of C-H singlet (C-H decoupled spectrum) as well as C-D triplet, as shown in an inset of Fig. 2. This proves that the ¹H signal at 7.23 ppm is not an impurity but the real signal of C-H group of enolic tautomer.

Signals at 7.33 *ppm* (s, 1H), 7.80 *ppm* (t, 1H), and 8.12 *ppm* (d, 2H), although very weak, indicate the existence of enol III form, but its concentration is negligible. In highly acidic media (pH < 0), enolization rate constant is very low [31], allowing the coexistence of two enol tautomers in solution.

HMBC spectrum (Fig. 3) provides important data about the major tautomeric form of 4PDA in CF₃COOD solution. Strong coupling between *ortho*-phenyl hydrogens and C_{δ} keto group carbon atom (circled signal) indicate that enol I is the major form of 4PDA in solution. If an enol form III was the predominant one, no coupling between *ortho*-hydrogens and C_{β} would be visible in HMBC spectrum. This confirms the predominance of enol I form of 4PDA in highly acidic solution.

Calculated ¹H NMR spectrum for diketo tautomer II has signals at 3.89 and 5.14 *ppm*, assigned to H atoms from > CH₂ group; corresponding C_{γ} atom has signal at 36.38 *ppm* in ¹³C spectrum. The asymmetric structure of the most stable geometry of 4PDA in a diketo form II results in the different chemical environment of two H atoms of a methylene group. As a consequence, two signals appeared in calculated ¹H NMR spectrum.

^b (d, J = 7.37 Hz, 2H)

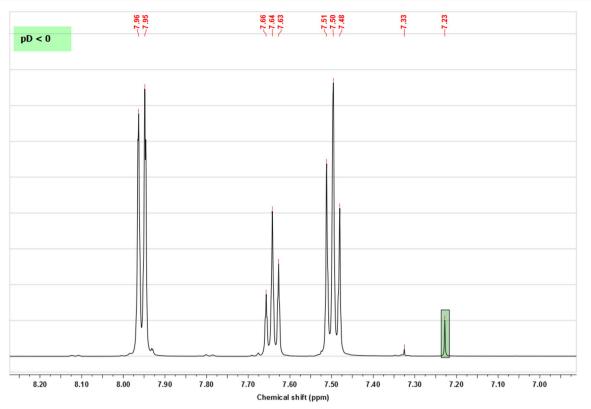


Fig. 1 ¹H NMR spectrum of 4PDA in CF₃COOD ($t = 25 \pm 1$ °C), where compound exists solely as an H₂A form. The signal of enolic, H γ atom is highlighted (green box)

It was mentioned that signals characteristic for the diketo tautomer were not observed in experimentally obtained NMR spectra of 4PDA in CF₃COOD. During routine characterization of some ADKs in aprotic solvents (CDCl₃ or DMSO-*d*₆), signals characteristic for diketo tautomer were present in ¹H and ¹³C NMR spectra. H atoms from > CH₂ group give one broad signal at 4.2 to 4.5 *ppm*, depending on substitution pattern on the phenyl ring, while in ¹³C NMR spectrum, the signal at ~ 50 *ppm* could be found [18–20].

Calculated NMR shifts of enolic hydrogens (H_{γ}) are different for two enol tautomers (Table 1). Thus, the low intensity signal at 7.33 *ppm* in experimentally obtained ¹H NMR spectrum (Fig. 1) could be ascribed to the second, less abundant, enol tautomer III.

¹H NMR spectra of 4PDA were also acquired in solutions with different acidity, within pD range of 1–10. As the acidity decreases, ¹H NMR spectrum becomes more complicated because the carboxylic group dissociates, leading to coexistence of H₂A and HA⁻ forms in solution. NMR spectrum of 4PDA in D₂O/CD₃COOD mixture (pD = 2.49) is shown in Fig. 4.

Two forms of 4PDA (H₂A and HA⁻) were present in this solution as is expected according to experimentally obtained pK_{a1} value (2.06) [20]. Separated doublets at 7.99 (H₂A) and 8.02 *ppm* (HA⁻) correspond to the *ortho*-H atoms. The molar ratio of H₂A to HA⁻ in a solution ($n_{H2A}/n_{HA-} = 0.78$) was calculated from corresponding peak areas, and was in good

agreement with the ratio $(n_{\rm H2A}/n_{\rm HA-} = 0.93)$ calculated using the Henderson-Hasselbalch equation. Two overlapped triplets at ~ 7.7 *ppm* correspond to *para*-H atoms of two ionization forms, while *meta*-H atoms (at ~ 7.6 *ppm*) are overlapped and could not be distinguished. The signal of H_{γ} atom was observed at 7.04 *ppm*, but the intensity was even smaller than in CF₃COOD, as the exchange rate between H_{γ} and D atom in D₂O/CF₃COOD is faster than in pure CF₃COOD. Lack of corresponding signals of enol III and diketo form II confirmed that the enol I is the dominant form of 4PDA in a solution.

NMR spectra were also recorded in deuterated acetate buffer, pD 4.81 (Fig. 5), where 4PDA, according to pK_{a2} value (7.56, [20]), is expected to be present in a pure HA⁻ form. No signals of diketo tautomer were observed. Signals of ortho-H (d, 7.99 ppm), para-H (t, 7.69 ppm), and meta-H (t, 7.58 ppm) are all doubled with meta-H being the least resolved, indicating the coexistence of another species in a solution. Although there is a possibility for a free rotation around C_{γ} - C_{δ} bond in enol I, intramolecular H-bond "locks" the pseudo 6membered ring, making the conformer/rotamer in which enolic oxygen on C_{β} and keto oxygen on C_{δ} are proximal to each other, a predominant one. Deprotonation of carboxyl group enables the formation of intramolecular hydrogen bond between carboxylate's oxygen and enolic –OH group on C_{β} , forming a pseudo 5-membered ring. This facilitates rotation around the $C_{\gamma}\text{-}C_{\delta}$ bond, so different conformers/rotamers, in

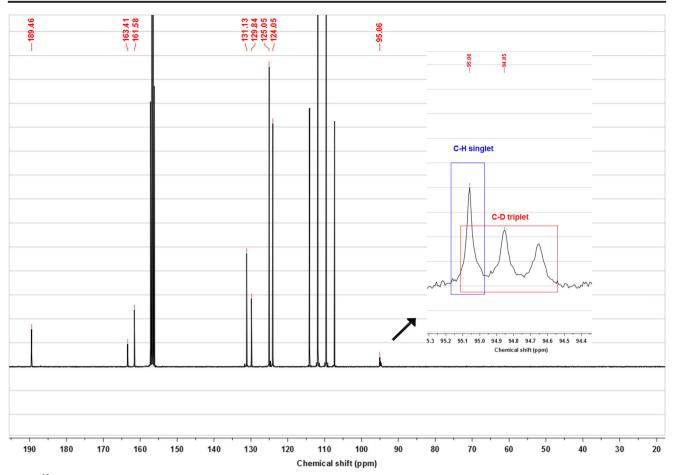


Fig. 2 ¹³C NMR spectrum of 4PDA in CF₃COOD; ($t = 25 \pm 1$ °C). The inset shows a magnified area around 95 ppm

which keto oxygen on C_{δ} and enolic oxygen on C_{β} are distal from each other, can exist in a solution. Therefore, we assume that the presence of two (stable) rotamers around C_{γ} - C_{δ} bond causes the duplication of signals of aromatic protons in ¹H NMR spectrum. Similar explanation is given in the literature to justify the complex ¹H NMR spectrum of ADK methyl esters sodium salts. The formation and coexistence of *Z*,*Z*and *E*,*Z*- isomers is described, as a consequence of rotation around C_{β} - C_{γ} and C_{γ} - C_{δ} bonds [33].

At pD 8.20, two forms of 4PDA (HA⁻ and A²⁻) are observed in a solution (Fig. 6), and the ratio of two forms was again in a good agreement with the one predicted from pK_a values.

In carbonate buffer at pD = 9.60 dianionic (A^{2–}) form should be the dominant species (Fig. 7). *Ortho*-H atoms appeared as a doublet at 7.72 *ppm*, and *meta-* and *para-*H atoms as overlapped triplets around 7.40 *ppm*. As π -electrons of dianion are delocalized over an entire keto-enol moiety, the spectral distinction between tautomers is not possible. Thus, two weak, but visible signals (t, δ = 7.57 *ppm* and d, δ = 7.82 *ppm*) indicated the presence of HA[–] with the abundance < 10%, which is in accordance with measured pD and pK_{a2} value. A summary of ¹H NMR shifts at all studied pD values, and calculated vs. predicted ratio of different protonation forms of 4PDA is given in Table 2.

Several observations important for understanding the solution chemistry of 4PDA should be stressed. The area under the ¹H NMR signal of enolic hydrogen atom (H_{γ}) is less than expected since it exchanges with the ²D from the solvent. With the increase of pD value, the intensity of H_{γ} decreases. This results in absence of H_{γ} signal in solutions with high pD values (8.20 and 9.60). As compound's, the carboxylic group dissociates the electron density of nucleus increases and enolic hydrogen atom becomes more shielded.

At pD 2.49, 8.20, and 9.60, 4PDA was found in solution as a mixture of two ionization forms, whose ratio of NMR peak areas correlates well with the ratio calculated from pK_a values (Table 2). When a mixture of two ionization forms is present in a solution, NMR signals of *ortho*-H atoms are resolved much better than the signals of *para*-atoms, while *meta*-H signals of two forms are overlapped. A transfer of negative charge from 4PDA (di)anion to aromatic ring is the most effective in *ortho*-position, because of the influence of both inductive and resonance effect. Since resonance effect is transferred only through *ortho*- and *para*-positions of the phenyl

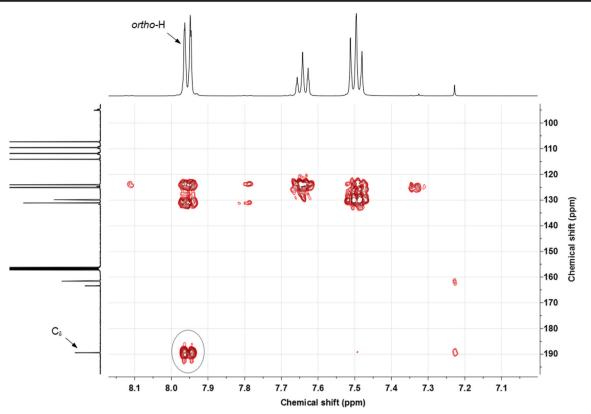


Fig. 3 HMBC NMR spectrum of 4PDA in CF₃COOD; $t = 25 \pm 1$ °C; I = 0.1 M (NaNO₃). The encircled cross-peak depicts the long-range correlation between *ortho*-phenyl hydrogens and C_{δ} atom

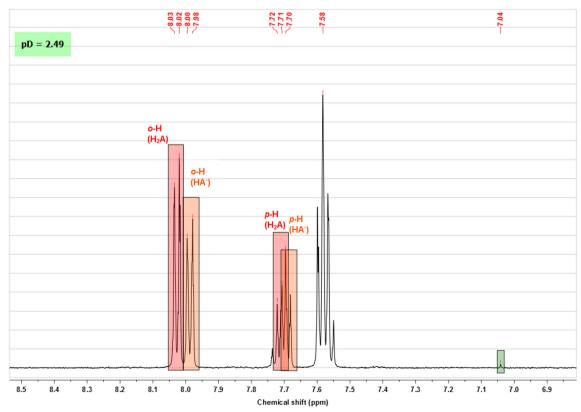


Fig. 4 A characteristic part of ¹H NMR spectrum of 4PDA in the D_2O/CD_3COOD mixture, pD = 2.49; $t = 25 \pm 1$ °C; I = 0.1 M (NaNO₃). Highlighted are signals of *ortho*- and *para*-hydrogens of molecular (red) and monoanionic (orange) form

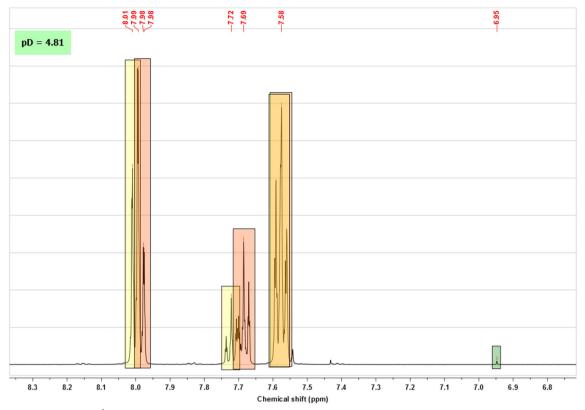


Fig. 5 A characteristic part of ¹H NMR spectrum of 4PDA in a deuterated acetate buffer, pD = 4.81; $t = 25 \pm 1$ °C; I = 0.1 M (NaNO₃). Highlighted (yellow and orange boxes) are signals of two forms, which could not be ascribed to different ionization forms (see the discussion below)

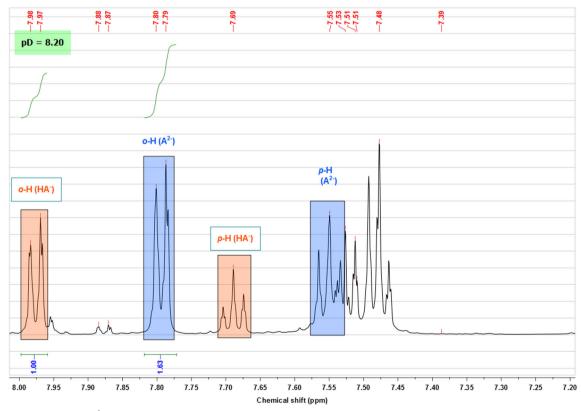


Fig. 6 A characteristic part of ¹H NMR spectrum of 4PDA in a carbonate buffer, pD = 8.20; $t = 25 \pm 1$ °C; I = 0.1 M (NaNO₃). Highlighted are signals of *ortho*- and *para*-hydrogens of monoanionic (orange) and dianionic (blue) form

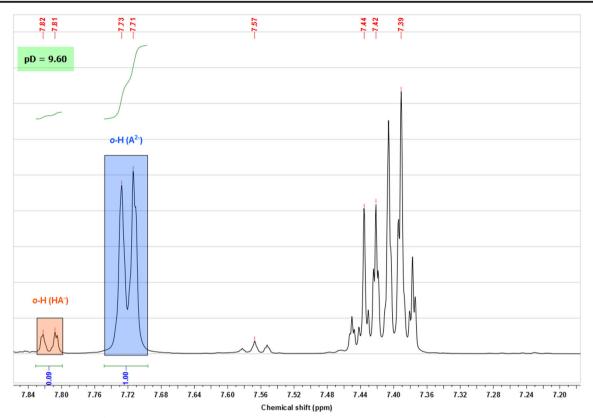


Fig. 7 A characteristic part of ¹H NMR spectrum of 4PDA in carbonate buffer, pD = 9.60; $t = 25 \pm 1$ °C; I = 0.1 M (NaNO₃). Highlighted are signals of *ortho*-hydrogens of monoanionic (orange) and dianionic (blue) form

ring, *para*-H atom is more influenced by a negative charge of dioxobutanoic moiety than *meta*-H atoms.

electron density of dioxobutanoic moiety is, to some extent, inductively transferred to aromatic ring.

Going from molecular to dianionic form, chemical shifts of all aromatic H atoms are moving upfield. It shows that the At pD 4.81, we hypothesized the coexistence of two stable rotamers in a solution, and the trend in spectral resolution of

Table 2 Summary of NMR shifts of 4PDA in buffered aqueous solutions. ¹H NMR shifts of H_2A , HA^- , and A^{2-} forms are shown in red, black, and blue, respectively

		δ (p	pm)		H ₂ A/HA ⁻ ratio calculated according to		HA^{-}/A^{2-} ratio calculated according to	
pD	Hγ	H ₂	H ₃	H_4	NMR spectra	p <i>K</i> _a values	NMR spectra	p <i>K</i> _a values
<0	7.23	7.96	7.50	7.64	Pure H ₂ A	Pure H ₂ A	/	/
2.49	7.04	<mark>8.02</mark> (7.99)	7.58	7.72 (7.70)	0.78	0.93	/	/
4.81	6.95	7.99	7.58	7.72 (7.69)	/	Pure HA ⁻	/	Pure HA ⁻
8.20	/	7.98 (7.80)	7.53 (7.48)	7.69 (7.55)	/	/	0.61	0.58
9.60	/	7.82 (7.72)	7.39	7.57 (7.44)	/	/	0.09	0.03

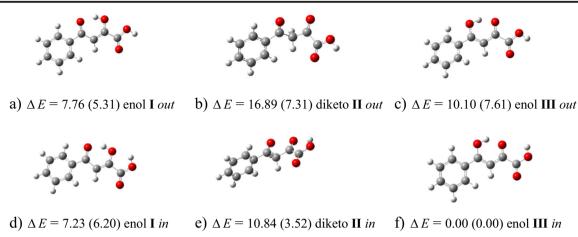


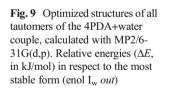
Fig. 8 Optimized structures of three 4PDA tautomers with carboxyl H *out* (**a**–**c**) and *in* orientations (**d**–**f**), calculated with MP2/6-31G(d,p). Relative energies (ΔE , in kJ/mol) *in vaccuo* and in implicit solvation model of water (values in brackets) with respect to enol III *in* tautomer (**f**)

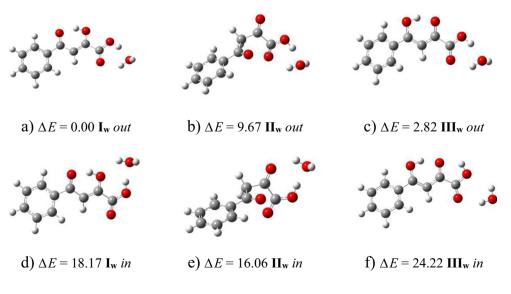
aromatic H atoms remained the same as when two ionization forms are present.

A computational study of the stability of 4PDA tautomeric forms

The stability of different tautomers of 4PDA in molecular form (H₂A) was investigated using quantum chemical calculations employing MP2 Hamiltonian, and 6-31G(d,p) basis set. For all three tautomers, forms with carboxyl hydrogen oriented toward surroundings (*out* orientation) and with carboxyl hydrogen oriented inside (*in* orientation) were considered. The inclusion of implicit solvation model (H₂O) led to the stabilization of all tautomeric forms studied. Such stabilization was larger for diketo than for enol tautomers of 4PDA. Larger dipole moments of diketo comparing to enol tautomers, calculated with implicit solvation model (ESM, Table S1), indicate a possibility for more favorable electrostatic interactions between polar solvent and diketo tautomers. Additional intramolecular hydrogen bond appears in the geometries with carboxyl hydrogen *in* when compared to *out* forms, and hence a possibility for the interaction of carboxyl hydrogen with a polar solvent is reduced. Although an implicit solvation model had larger influence on forms with carboxyl hydrogen oriented toward surroundings (*out* orientation), the intramolecular hydrogen bonding made forms with carboxyl hydrogen oriented *in* more stable, both in calculations with and without implicit solvent model applied. Geometries of the lowest energy forms of 4PDA with carboxyl hydrogen in both orientations are given in Fig. 8 a–f. Relative energies are given in respect to the apparently most stable tautomer (enol III *in*), for calculations in a vacuum and in implicit solvation model.

Energy differences between two enol tautomers are 7.23 kJ/mol in vacuum and 6.20 kJ/mol in implicit solvent model, for *in* orientation of carboxyl hydrogen (enol I *in* vs. enol III *in*). Diketo forms were less stable, but such difference was significantly reduced upon the inclusion of implicit





solvent in calculations (from 10.84 to 3.52 kJ/mol for diketo II *in*). Comparing three tautomers having carboxyl H *out* orientation, the most stable form was enol I. Energy differences between enol I and enol III are 2.34 kJ/mol *in vaccuo* and 2.30 kJ/mol when implicit solvation was accounted. Enol I is 9.13 kJ/mol more stable than diketo II form *in vaccuo*, and only 2.00 kJ/mol in implicit water model.

Even though calculations suggested that forms with *in* orientation of carboxylic H were more stable than corresponding *out* forms, it was reasonable to assume the existence of stable *out* forms in a real situation, i.e., in a protic solvent like water. It is well known that a PCM model is not good for simulating solvents with hydrogen-bonding properties. The existence of tautomer III *in* which was predicted as the most stable form could be questioned, since no experimental data point to the existence of tautomer III in detectable amount for ADKs, or structurally similar compounds. Literature data on quantum chemical calculations of L-731,988, aryldiketo acid derivative having *N*-benzyl substituted pyrrole as aroyl moiety, describe also an analog of enol III as the most stable one when solvation effects were accounted using the PCM model of water [17].

As implicit solvation model accounts for dipolar interactions, influencing mainly C-heteroatom and heteroatom-H bonds polarization, we added one explicit water molecule in the proximity of carboxyl group, and optimized geometry of such system for all six forms. Results are shown in Fig. 9.

The enol I with carboxyl H in *out* orientation + water couple (enol I_w) appeared as the most stable of all six forms, followed by enol III_w ($\Delta E = 2.82$ kJ/mol) and diketo II_w ($\Delta E = 9.67$ kJ/mol) forms. All tautomers with carboxyl H *in* orientation (I_w *in*, III_w *in*, and I_w *in*) appeared significantly less stable compared to *out* counterparts. Thus, an inclusion of one explicit water molecule in the model, which allowed carboxyl H to establish its (intermolecular) hydrogen-bond-donating ability, provided results in much better agreement with experiments.

Conclusions

Aryldiketo acids complexation ability with divalent metal ions, that could be responsible for their mode of biological action, and the hydrolytic C–C bond cleavage by β ketolases depend on predominant tautomeric form. To the best of our knowledge, no experimental data on keto-enol tautomerism of ADKs in aqueous solution with pH \leq 5.5 was published so far. In a highly acidic medium, 4-phenyl-2,4dioxobutanoic acid (4PDA) predominantly exists as an enolic form with keto group closer to the phenyl ring. At higher pH values, where 4PDA exists as a mixture of two species with different ionization states, NMR pattern becomes more complex. The ratio of two forms is in a good agreement with the ratio predicted from the Henderson-Hasselbalch equation. An exception was found in ¹H NMR spectrum of 4PDA monoanion where signals are doubled, and the coexistence of two stable rotamers is hypothesized. Ab initio MP2/6-31G(d,p) method accurately predicted the relative stability of tautomers only when explicit water molecule was included in calculations.

Our combined experimental and computational evidences strongly suggest that enol I form is most prominent form in biologically relevant environments that these molecules can be exposed to. These observations could be taken into account in future efforts to develop drug-like molecules based on this class of compounds. Furthermore, a similar approach can be used to assess the prevalence of tautomers for other classes of molecules with diketo moiety. Findings from this study may be important for further development of this type of compounds as drugs, since absorption, distribution, and other pharmacokinetic properties depend on a tautomeric form of a compound.

Funding Information Ministry of Education, Science, and Technological Development of Serbia supported this work, Grant No. 172035.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Wai JS, Egbertson MS, Payne LS, Fisher TE, Embrey MW, Tran LO, Melamed JY, Langford HM, Guare Jr JP, Zhuang L, Grey VE, Vacca JP, Holloway MK, Naylor-Olsen AM, Hazuda DJ, Felock PJ, Wolfe AL, Stillmock KA, Schleif WA, Gabryelski LJ, Young SD (2000). J Med Chem 43:4923–4926
- Hazuda DJ, Felock P, Witmer M, Wolfe A, Stillmock K, Grobler JA, Espeseth A, Gabryelski L, Schleif W, Blau C, Miller MD (2000). Science 287:646–650
- Tan JJ, Liu C, Sun HX, Cong JX, Hu ML, Wang XC, Liang JX (2012). Mini Rev Med Chem 12:875–889
- Cuzzucoli Crucitti G, Métifiot M, Pescatori L, Messore A, Madia VN, Pupo G, Saccoliti F, Scipione L, Tortorella S, Esposito F, Corona A, Cadeddu M, Marchand C, Pommier Y, Tramontano E, Costi R, Di Santo R (2015). J Med Chem 58:1915–1928
- Tomassini J, Selnick H, Davies ME, Armstrong ME, Baldwin J, Bourgeois M, Hastings J, Hazuda D, Lewis J, McClements W (1994). Antimicrob Agents Chemother 38:2827–2837
- Drakulić BJ, Stavri M, Gibbons S, Žižak ŽS, Verbić TŽ, Juranić IO, Zloh M (2009). ChemMedChem 4:1971–1975
- 7. Milletti F, Vulpetti A (2010). J Chem Inf Model 50:1062-1074
- Sechi M, Bacchi A, Carcelli M, Compari C, Duce E, Fisicaro E, Rogolino D, Gates P, Derudas M, Al-Mawsawi LQ, Neamati N (2006). J Med Chem 49:4248–4260
- Billamboz M, Bailly F, Barreca ML, De Luca L, Mouscadet JF, Calmels C, Andréola ML, Witvrouw M, Christ F, Debyser Z, Cotelle P (2008). J Med Chem 51:7717–7730
- 10. Koz'minykh VO, Koz'minykh EN (2004). Pharm Chem J 38:67-77
- 11. Reguera J, Weber F, Cusack S (2010). PLoS Pathog 6:e1001101

- 12. DuBois RM, Slavish PJ, Baughman BM, Yun M-K, Bao J, Webby RJ, Webb TR, White SW (2012). PLoS Pathog 8:e1002830
- Krieger IV, Freundlich JS, Gawandi VB, Roberts JP, Gawandi VB, Sun Q, Owen JL, Fraile MT, Huss SI, Lavandera JL, Ioerger TR, Sacchettini JC (2012). Chem Biol 19:1556–1567
- 14. Zhu W, Zhang Y, Sinko W, Hensler ME, Olson J, Molohon KJ, Lindert S, Cao R, Li K, Wang K, Wang Y, Liu YL, Sankovsky A, de Oliveira CA, Mitchell DA, Nizet V, McCammon JA, Oldfield E (2013). Proc Natl Acad Sci U S A 110:123–128
- Brecker L, Pogorevc M, Griengl H, Steiner W, Kappe T, Ribbons DW (1999). New J Chem 23:437–446
- Sechi M, Derudas M, Dallocchio R, Dessì A, Bacchi A, Sannia L, Carta F, Palomba M, Ragab O, Chan C, Shoemaker R, Sei S, Dayam R, Neamati N (2004). J Med Chem 47:5298–5310
- 17. Huang M, Richards WG, Grant GH (2005). J Phys Chem A 109: 5198–5202
- Cvijetić IN, Verbić TŽ, Drakulić BJ, Stanković DM, Juranić IO, Manojlović DD, Zloh M (2017). J Serb Chem Soc 82:303–316
- Verbić TŽ, Drakulić BJ, Zloh M, Juranić IO (2008). Lett Org Chem 5:692–699
- Verbić TŽ, Drakulić BJ, Zloh MF, Pecelj JR, Popović GV, Juranić IO (2007). J Serb Chem Soc 72:1201–1216
- Kojić M, Petković M, Etinski M (2016). Phys Chem Chem Phys 18:22168–22178
- Kojić M, Petković M, Etinski M (2016). J Serbian Chem Soc 81: 1393–1406
- Belova NV, Oberhammer H, Girichev GV (2004). J Phys Chem A 108:3593–3597

- 24. Moriyasu M, Kato A, Hashimoto Y (1986). J Chem Soc Perkin Trans 2:515–520
- 25. Yamabe S, Tsuchida N, Miyajima K (2004). J Phys Chem A 108: 2750–2757
- 26. Sigalov MV (2015). J Phys Chem A 119:1404-1414
- 27. Glasoe PK, Long FA (1960). J Phys Chem 64:188-189
- 28. Wolinski K, Hinton JF, Pulay P (1990). J Am Chem Soc 112:8251– 8260
- 29. MOPAC2016, Stewart JJP (2016) Stewart Computational Chemistry, Colorado Springs, CO, USA, http://OpenMOPAC.net
- 30 Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Mennucci B, Petersson GA, Nakatsuji H, Caricato M, Li X, Hratchian HP, Izmaylov AF, Bloino J, Zheng G, Sonnenberg JL, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery JA, Peralta JE, Ogliaro F, Bearpark M, Heyd JJ, Brothers E, Kudin KN, Staroverov VN, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant JC, Iyengar SS, Tomasi J, Cossi M, Rega N, Millam JM, Klene M, Knox JE, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Martin RL, Morokuma K, Zakrzewski VG, Voth GA, Salvador P, Dannenberg JJ, Dapprich S, Daniels AD, Farkas Ö, Foresman JB, Ortiz JV, Cioslowski J, Fox DJ (2009) Gaussian 09, Revis. B.01. Gaussian, Inc., Wallingford
- 31. Guthrie JP (1972). J Am Chem Soc 94:7020-7024
- Maurin C, Bailly F, Buisine E, Vezin H, Mbemba G, Mouscadet JF, Cotelle P (2004). J Med Chem 47:5583–5586
- 33. Maurin C, Bailly F, Cotelle P (2004). Tetrahedron 60:6479-6486